In Silico Elucidating the Common Gene-Regulatory-Network(S) Of Accelerated Cell Death (Acd11) from Arabidopsis

Thaliana and Its Orthologs

Shahid Ali¹, Humayun², Mushtaq Ali³, Muhammad Ali⁴,

Fazal Raziq⁵, Saadat Khan⁶

ABSTRACT

The Arabidopsis accelerated cell death 11 (acd11) is a genetic model for studying immune response activation and localized cellular suicide that prevent pathogen spread during plant infection. In this research article we predicted the common gene-regulatory-network of accelerated cell death (acd11) from Arabidopsis thaliana and its orthologs with the help of well-known bioinformatics tools. The most common orthologs were extracted from Phytzome and PLAZA on the basis of protein sequence similarity using BLASTp. Developmental expression analysis from Genevestigator concluded the acd11 is highly expressed at last developmental stage and co-expressed with Autophagy related and Disease resistant genes. PPI from STRING network confirm that its interacting partners are EDS1 and LAZ1 both of which play an important role in disease resistant and programmed cell death respectively. Protein disorder from DisEMBL confirm its diverse functioning properties and non-stable structure. Ligand binding study from RaptorX predicted that residues in the Active site pocket Asparagine and Histidine.

Keywords: Arabidopsis accelerated cell death 11, acd11, BLASTp

INTRODUCTION

Due to the hypersensitive (HR) response in plants, which causes PCD, the plant host resistance (R) protein identifies pathogen avirulence component. In the model plant Arabidopsis, recessive acd11 is a lesion mimic mutant that activates HR-like PCD in the absence of pathogen infection. Before the plant flowers, Acd11 starts to degrade the chlorophyll in the two-leaf stage, consuming the entire plant (Brodersen *et al.*, 2002). Sphingolipid transfer proteins that help sphingosine transfer across membranes in vitro

Manuscript Received: Oct 12, 2022 Manuscript Accepted: 29 Dec, 2022

¹ Assistant Professor, Govt. Postgraduate College Swabi

Corresponding Author's Email: profshahid.ali@gmail.com

² Assistant Professor, Govt. Degree College Yar Hussain, Swabi

³ Assistant Professor, Govt. Postgraduate College, Swabi

⁴ Assistant Professor, GDC Toru, Mardan

⁵ Assistant Professor, GPGC Mardan

⁶ Assistant Professor, Govt. Commerce College, Peshawar

are homologous to ACD11 (Brodersen *et al.*, 2002). ACD11 has similarity to GLTP, a protein that facilitates the transfer of glycoshingolipids. Although mammalian GLTP is structurally and biophysically well described, its biological function is unknown (Brown and Mattjus, 2007). The positively charged triad's clustered Lys/Arg residue in ACD11, which is best positioned to bind phosphate, explains why the protein is unable to bind sugar head groups or transfer glycolipids (Peterson *et al.*, 2008).

ACD11 in yeast forms interactions with PRA7 and PRA8, while VAP27-1 has a stronger interaction with an ACD11 homolog rather than the ACD11 protein itself. While its interactors have varying degrees of membrane association, ACD11 is mostly a cytosolic protein. ACD11, in conjunction with the proteins PRA and VAP, may control membrane trafficking (Peterson *et al.*, 2009).

To regulate programmed cell death (PCD), it is essential to maintain a dynamic balance between Cer and C1P as per the studies by Chen *et al.* (2009), Pata *et al.* (2010), and Berkey *et al.* (2012). The deficiency of *acd11* results in altered C1P levels and increased phyto-cer levels, indicating a functional connection between *acd11* expression and sphingolipid metabolic regulation in plants. Despite the low sequence similarity among ACD11 and other GLTP homologs, such as human CPTP, Simanshu *et al.* (2014) have demonstrated the existence of conserved structural similarity between Arabidopsis ACD11, human GLTP, and human CPTP.

OBJECTIVES

- 1. Identification and extraction of orthologs of ACD11 from *Arabidopsis thaliana* among plant species.
- 2. Screen for common interacting protein with ACD11 and its orhtologs to analyze and deduce the gene regulatory network of ACD11.
- 3. Prediction of 3D structure and interacting residue of proteins by determining its interface residue, intrinsic protein disorder.

LITERATURE REVIEW

Simanshu *et al.* (2014) have demonstrated that ACD11, encoded by the *acd11* gene, is a lipid transfer protein that promotes intermembrane transfer of sphingosine and sphingomyelin but not Cer or glycolceremides. *Acd11* is a lesion mimic mutant in the model plant *Arabidopsis*, and in its recessive form, it triggers HR-like PCD even without pathogen infection. At the two-leaf stage, *Acd11* induces chlorophyll degradation, which eventually engulfs the entire plant before flowering. Despite being similar to three known sphingolipid transfer proteins, ACD11 has been demonstrated to aid the transfer of sphingosine between membranes in laboratory experiments.

The discovery of lesion mimic mutants (LMM), such as the hypersensitive response (HR), which is associated with pathogen resistance, has proven to be a useful tool in studying programmed cell death (PCD) pathways in plants (Lorrain *et al.*, 2003).

The incompatibility reaction, a cell death response that takes place when cells of different genotypes fuse in filamentous fungi, is governed by a group of loci called *het* loci (heterokaryon incompatibility loci). In *Podospora anserina*, the genes upregulated during this cell death response are involved in a process similar to autophagy, and ACD11 homolog, called HET-C protein, is implicated in vegetative incompatibility reaction in the fungus (Pinan-Lucarre *et al.*, 2003).

Plants rely on programmed cell death (PCD) as an essential response mechanism to various internal and environmental signals to achieve proper development and survival. Unlike animal apoptosis, PCD in plants is characterized by the absence of engulfment by neighboring cells. PCD occurs in plants during development, virulent infections, and the hypersensitive response (HR) brought about by avirulent stress effectors, as noted by Lam and colleagues in 2004.

The plant surveillance system has evolved to recognise pathogen-secreted chemicals in order to initiate a defense response when plant pcd coexists with disease resistance, an occurrence known as the hypersensitive response. These released compounds function as virulence factors in plants lacking genetic disease resistance, acting through mostly unidentified methods. If the function of ACD11 is lost, the result is a lethal condition. The recessive homozygous mutant of *acd11* grow normally until they develop 2-4 leaves. After that, they exhibit accelerated cell death similar to that observed in lesion mimic mutant, which is a plant with spontaneous lesions on the leaves (Greenberg *et al.*, 2004).

Broderson *et al.* (2005) have reported that salicylic acid (SA) levels increase in response to infections that induce programmed cell death (PCD), and salicylate hydroxylase, which is encoded by the bacterial nahG gene, can prevent PCD development. SA is believed to play a role in the initiation of PCD linked to pathogen defense responses. PCD in acd11 is reliant on the plant hormone SA and can be hindered by expressing a bacterial SA hydroxylase (nahG) or through mutation in PAD4 and EDS1.

Airene et al. (2006), Brown and Mattjus (2007), and Peterson et al. (2008) have suggested that ACD11 has a GLTP fold and acts as a glycolipid transfer protein, based on its predicted structural homology model. Meanwhile, Hofius et al. (2007) reported that programmed cell death (PCD) during the hypersensitive response (HR) in plants is triggered by recognition of pathogen avirulence factors by the plant host resistance (R) protein.

According to research by various authors, including Brown and Mattjus (2007), GLTP and FAPP2 are homologs of ACD11 that are involved in the transfer of glycosphingolipids between membranes, although the exact role of GLTP remains unknown. ACD11 and ACD5 are involved in regulating the balance of Cer and C1P levels, which control PCD related to HR. Additionally, ACD11 may play a role in regulating membrane trafficking through interactions with PRA and VAP proteins, despite its inability to transfer glycolipids due to the Lys/Arg residue cluster. Studies also suggest that ACD11 expression is involved in regulating sphingolipid metabolism in plants, and its deficiency can lead to inappropriate HR activation by LAZ5 (Chen *et al.*, 2009; Pata *et al.*, 2010; Berkey *et al.*, 2012; Peterson *et al.*, 2008, 2009; Wang *et al.*, 2008; Palma *et al.*, 2010).

LAZ1, a DFU300 transmembrane protein, serves as a regulator of PCD associated with HR and suppresses cell death in acd11 and certain types of HR cell death (Malinovsky et al., 2010). Although ACD11 and other GLTP homologs, including human CPTP, have low sequence homology, crystallographic data has established their conserved structural homology (Simanshu et al., 2014).

RESEARCH METHODOLOGY

The research methodology of this research article is described in the following subsections.

Orthologous genes of ACD11 that is identifie and selected on the basis of protein sequence similarity from PLAZA, PHYTOZOME (Goodstein *et al.*, 2012) and NCBI Databases.

Protein sequences of the orthologous genes will be retrieved from Phytozome for further analysis.

Gene regulatory network will be predicted on the basis of tissue or conditional specific co-expressional behavior of genes *acd11*, using expression browser Databases such as eFP and tools like Planex (Yimet al., 2013), PlaNet (Mutwil *et al.*, 2011) and Genevestigator (Zimmerman *et al.*, 2004),

PPI network of ACD11 and orthologous genes will be retrieved from STRING (Szklarczyk *et al.*, 2011) to predict common interacting protein. 3D structure of all interacting protein will be predicted from PHYRE2 and HHPred.

DisEMBL is a useful method for predicting protein structure disorder, and RaptorX is a web-based tool used for determining the functional role of proteins by generating reliable three-dimensional atomic models of proteins. These tools are important for target selection and constructing biochemical studies, particularly in genomics and structural biology projects. In addition, protein sequences and structures can be utilized to forecast residues that participate in ligand binding sites (LBS) (Linding and Jenssen *et al.*, 2003; Wang and Li *et al.*, 2016).

RESULT AND DISCUSSIONS

Identification of orthologous for the reference Protein ACD11

For the reference gene ACD 11 of Arabdosis Thaliana were retrieved and identified through MSA BLAST tool form NCBI and Phytozome on the basis of protein structure similarity. From the following retrieved data of orthologues from databases, further selection was made on the basis of their availability for protein interaction and co-expression analysis.

Table 1. Representing ACD 11 (AT2G34690) orthologous genes with GeneInfoIdentifiers from different plant species Data obtained from PLAZ and PhytozomeBLASTp search.

GENE ID PLAZA	GENE ID PHYTOZOME	ORGANISM	PROTEI N LEGTH	E- VALU E	BIT SCOR E	% IDENTIT Y
CRU_004G1584 0	Carubv10023844 m.g	C.rubella	207	4e-140	400	92.75
TC0002G01160	Thecc1EG006094	T. cacao	207	5e-108	316	72.46
GR08G04690	Gorai.008G04690 0	G. raimondii	207	2e-106	312	71.98
MD10G020560	MDP0000162581	M.domestica	207	2e-103	304	71.01
RC28617G00100	28617.t000010	R. communis	214	2e-103	305	69.16
PPE_004G08510	Prupe.4G089300	Prunus persica	201	5e-102	302	70.65
FV3G15140	1	Fragaria	201	1e-99	295	71.64

Ali, Humayun, Ali, Ali, Raziq, Khan

	gene28951-v1.0-	vesca				
SL02G070250	hybrid		205	1e-96	287	68.78
	Solyc02g070250.2	S.lycopersicu m				
ZM01G55250			209	1e-96	289	67.46
	GRMZM2G0588	Zea mavs				
OS03G57140	12		209	9e-93	277	61.72
		Orvza Sativa				
CS00010G00480	0	j	171	9e-87	262	74.27
MT2G029520	1g027852m.g	Citrus cinensis	209	5e-86	260	61.24
PT06G05180	Medtr2g030530	M.truncata	171	5e-83	253	71.35
	Potri.006G05180 0	P.trichocarpa				
GLYMA15G134 70	GM.15G13470	Glycine max	209	7e-99	293	70.05
	l	l	I			

The plant species selected for further study have almost the same number of Amino acids in polypepetide chain with variations between 214 aa to 171 aa. The selection of species cover both dicots (S.lycopersicum, M.truncata, P.trichocarpa, R.cummunis) and monocots (Zea mays and Oryza sativa), for predication their protein structure, function and interactions to validate the reference gene and protein regulatory network analysis.

Extraction of protein sequences of ACD11 orthologs

Protein sequences in FASTA format of the orthologues gene were retrieved from the Phytozome database for further analysis of protein structure prediction, ligand binding, protein interaction, protein disorder all based on the sequence of proteins.

Predicting developmental stages co-expression of ACD11 and its orthologues

The expression of genes was examined during plant development and at the anatomical level by accessing the Affymetrix array database using the Genevestigator response viewer. Microarray data from the Arabidopsis Gene Chip platform was obtained and analyzed by querying gene member IDs. The database contains microarray data from only the wild-type background, and the expression level was assessed in the late stages of plant development, including senescence. ACD11 loss-of-function mutants exhibit accelerated cell death similar to that observed in lesion mimic mutants, and the protein may play a functional role during plant maturation and senescence (Greenberg et al., 2004).

The co-expressed gene were retrieved form the data for the refence gene as well as of their orthologues.



Figure 1 Developmental stage-dependent expression analysis of the reference gene and its orthologues, in most cases the genes expression is at highest level in the last

developmental stages of plant. (a) Arabidopsis thaliana (b) Oryza staiva (c) Zea mayz (d) Lycopersicum esculentum (e) Medicago truncatula (f) Glycine max

Co-expression were observed under the stress conditions (Perturbations) by default with a 50 limit of expressed genes. Positive correlation of the expressed genes under the same conditions were analyzed Pearson correlation through Genevistiagtor affymytrix biochips. All genes of the orthologues were put to the same conditions to confirm validation of the co-expressed genes with ACD11 of Arabidopsis thaliana. As it is evident from the result that stress and autophagy related proteins are highly co-expressed in the reference gene as well as in its orthologues, which reflects a strong association of these proteins in programmed cell death regulation of the plant. The programmed cell death observed in *acd11* is controlled by the hormone salicylic acid (SA) and can be inhibited by the expression of a bacterial SA hydroxylase (nahG) or by mutations in PAD4 and EDS1, as reported by Broderson *et al.* (2005) and Feys *et al.* (2001).



Figure 2: Conditional stress co-expression analysis of reference gene ACD11 using Genevestigator co-expression tool.

 Table 2: Co-expressed genes of the reference gene and its orthologues under stress conditions using Genevestigator tool.

Protein	Arabidopsi s Thaliana	Glycine max	S.lycopersicum	Oryza sativa	Zea mays
Syntaxin	AT4G177 30	GLYMA01G007 60	SOLYC05G0130 50	OS03G01374 00	GRMZM5G8067 84
	AT4G321 50	GLYMA14G021 20	SOLYC01G1008 20		
	AT1G266	GLYMA09G138	SOLYC03G1220		

	70	40	90		
	AT3G606 00	GLYMA16G082 00	SOLYC02G0329 30		
	AT1G162 40	GLYMA02G422 80			
	AT2G369 00				
UQ Ligase	AT4G098 30	GLYMA10G431 20	SOLYC04G0808 10	OS08G01709 00	GRMZM2G1714 30
	AT2G360 60	GLYMA06G023 90		OS11G04836 00	
	AT5G324 40			OS08G05403 00	
Zinc finger	AT5G164 70	GLYMA14G167 17	SOLYC03G0633 20	OS07G01672 00	GRMZM2G0492 96
		GLYMA12G132 70	SOLYC09G0084 70	OS06G06972 00	GRMZM2G0000 14
		GLYMA10G430 90			
		GLYMA07G121 70			
Stress related	AT1G370 0	GLYMA10G283 00	SOLYC09G0903 70	OS07G05894 00	GRMZM2G1069 60
protein		GLYMA03G410 30	SOLYC01G0804 10	OS05G01227 00	
			SOLYC09G0726		

			70		
			SOLYC01G1046 00		
			SOLYC07G0448 50		
Autophag y related protein	AT2G459 80		SOLYC06G0341 60 SOLYC03G0967 90	OS03G01374 00	GRMZM2G0565 72 GRMZM2G1714 35
			00		
Ras rekated protein		GLYMA09G019 50	SOLYC06G0652 30		
F		GLYMA05G325 20	SOLYC01G1047 00		

Protein-protein interaction network(s) and 3D structure prediction

The PPI network of ACD11 and its orthologous genes was obtained from the STRING database to predict its interactions with other proteins. The analysis revealed ACD11's interaction with EDS1, LSD1, LAZ1, LCB, and FMO. LAZ1, which belongs to the DFU300 transmembrane protein family, regulates plant PCD associated with HR and is involved in acd11 cell death. Loss of LAZ1 function suppresses cell death in acd11 and certain types of HR cell death. The STRING database is a valuable tool for predicting protein interactions and understanding their functional impact (Malinovsky *et al.*, 2010).



Figure 3: PPI of the reference gene and orthologues retrieved from STRING database network. (a) Arabidopsis Thaliana (b) Zea mayz (c) Lycopersicum esculentum (d) Oryza sativa.

• •	,					
<u>≜node1</u>	node2	node1 accession	node2 accession	node1 annotation	node2 annotation	score
ACD11	ACD5	AT2G34690.1	AT5G51290.1	ACCELERATED CELL DEATH 11	ACCELERATED CELL DEATH 5	0.902
ACD11	AT1G14340	AT2G34690.1	AT1G14340.1	ACCELERATED CELL DEATH 11	RNA recognition motif-containing	0.800
ACD11	AT1G23070	AT2G34690.1	AT1G23070.1	ACCELERATED CELL DEATH 11	uncharacterized protein	0.588
ACD11	AT1G77220	AT2G34690.1	AT1G77220.1	ACCELERATED CELL DEATH 11	uncharacterized protein	0.579
ACD11	AT5G45230	AT2G34690.1	AT5G45230.1	ACCELERATED CELL DEATH 11	TIR-NBS-LRR class disease resist	0.580
ACD11	BPA1	AT2G34690.1	AT5G16840.2	ACCELERATED CELL DEATH 11	binding partner of acd111	0.938
ACD11	EDS1	AT2G34690.1	AT3G48090.1	ACCELERATED CELL DEATH 11	enhanced disease susceptibility 1	0.753
ACD11	EDS16	AT2G34690.1	AT1G74710.2	ACCELERATED CELL DEATH 11	Isochorismate synthase 1; Involve	0.641
ACD11	FMO	AT2G34690.1	AT1G12200.1	ACCELERATED CELL DEATH 11	flavin monooxygenase; Catalyzes	0.597
ACD11	FM01	AT2G34690.1	AT1G19250.1	ACCELERATED CELL DEATH 11	flavin-dependent monooxygenase	0.597
ACD11	LAZ1	AT2G34690.1	AT4G38360.2	ACCELERATED CELL DEATH 11	LAZARUS 1	0.721
ACD11	LAZ5	AT2G34690.1	AT5G44870.1	ACCELERATED CELL DEATH 11	LAZARUS 5	0.759
ACD11	LCB2	AT2G34690.1	AT5G23670.1	ACCELERATED CELL DEATH 11	long chain base2; Serine palmitoyl	0.758
ACD11	LSD1	AT2G34690.1	AT4G20380.8	ACCELERATED CELL DEATH 11	zinc finger protein LSD1	0.711
ACD11	PRA7	AT2G34690.1	AT1G55190.1	ACCELERATED CELL DEATH 11	PRA1 family protein F2; May be in	0.894
ACD11	PRA8	AT2G34690.1	AT3G13720.1	ACCELERATED CELL DEATH 11	PRA1 family protein F3; May be in	0.939
ACD11	SAG13	AT2G34690.1	AT2G29350.1	ACCELERATED CELL DEATH 11	senescence-associated gene 13	0.645
ACD11	SBH1	AT2G34690.1	AT1G69640.1	ACCELERATED CELL DEATH 11	sphingoid base hydroxylase 1; Inv	0.645
ACD11	SBH2	AT2G34690.1	AT1G14290.1	ACCELERATED CELL DEATH 11	sphingoid base hydroxylase 2; lnv	0.621
ACD11	VAP27-1	AT2G34690.1	AT3G60600.1	ACCELERATED CELL DEATH 11	vesicle associated protein; May pl	0.861

	A.thalian a	S.lycopersicu m	G.max	P.trichocarpa	O.sativa	Zea mays
EDS	At3g480	solyc06g0712	glyma04g348	Poptr_0015s08	Loc_os09g224	grmzm2g0457
1	90	80	00	100	50	16
LSD	at4g2038	solyc02g0697	glyma05g304	Poptr_0011s15	Loc_os12g417	grmzm2g1146
1	0	20	90	810	00	13
LAZ	at4g3836	solyc07g0543	glyma06g003	10Poptr_0009s	Loc_os06g511	grmzm2g0004
1	0	20	00	167	00	27
LCB	at5g2367	solyc06g0723	glyma02g072	Poptr_0012s10	Loc_os01g703	grmzm2g0102
2	0	20	50	630	80	02
FM	at1g1220	solyc07g0424	glyma17g051	Poptr_0001s36	Loc_os03g084	grmzm2g0004
O	0	30	60	280	10	27

Table 3. Protein interacting partners of ACD11 and its orthologues

Phyre2 was used to predict the 3D structure of the reference protein and its orthologues, and HHPred was used to validate the structure by comparing the secondary structure of the reference protein to that of its orthologues. Although ACD11 has low sequence homology with other GLTP homologs, crystallographic data has shown conserved structural homology between Arabidopsis ACD11, human GLTP, and human CPTP. The 3D model for ACD11 was retrieved from the PDB database with ID 4NT1.



Figure 4: 3D Structure of the ACD11 retrieved from Phyre2

Prediction of ACD11 protein disorder and ligand binding residue and its characterization

It is noted that DisEMBL is a web-based tool used to study and predict Intrinsic Disorder Proteins (IDPs), and that understanding protein function and folding pathways is important for protein disorder. IDPs are thought to become ordered only when bound to another molecule or when biochemical environment changes. Loops/coils are not necessarily disordered, but protein disorder is only found within loops. Disordered regions in a protein's flexibility facilitate different conformational possibilities and are therefore important for functionality as these regions may contain functional sites or linear motifs. The functional diversity of the first and middle segments of the protein structure is revealed by Disorder by Hot-loops and Loop-coils.

S.NO	PROTEIN	RESIDUE POSITION	RESIDUES
1	A.thaliana	1-9, 92-107, 121- 131, 199-206	MADSEADKP, RKAGSHTRNLLRVKRG, <mark>SEGDNSLKDPA</mark> , SKQLGIDW
2	B.rapa	1-19 , 32-47, 62-73	MDYVAKVEDLAKASSSVST, CVRKAGSHTRNLLRVK, ASEGDNSLKDPA

 Table 4. Protein disorder by Hot-loop

3	S.lycopersicum	1-11, 99-112, 126- 136, 201-210	MANHVAEEKPL, AGSHTRNLLRVKRG, <mark>SEGNSLKDPAS</mark> , FTSRDLGTDW
4	G.max	1-8, 92-111, 124- 132	MTEGNGDK, NTVRKGGSHTRNLLRVKRGL, TEGNSLRDP
5	V.vinifera	86-104, <u>120-127</u>	IEGNCVRKAGSNSRNLLRV, ASDDNSLR
6	O.sativa	1-10, 77-108, 183- 192	<mark>MGSSDGDKPL</mark> , ISKLPEMVELDIQKGTVRQAGSHT RNMLRVKR, FIRASGPVIL
7	Z.mays	1-14, 22-59, 126- 161, 233-258	MNALKHRKARQHQ, QKNKKGEGKNSRLLPRACQ HPRSASSPKAAMGSSQAD, KSISTLPSMVERDIQ TDTVRKPGSHTRNLLRVKRGI, NFVRSSAP VICYVDDLFTSRNLGIDW
8	P.trichocarpa	1-10 , 73-107	<mark>MGDLETEKPL</mark> , KSIGTLQSVLDKDVERNS VRKGGSHSRNLLRVKRG

Table 5. Protein disorder by Loop/coil

S.N O	PROTEIN	RESIDUE POSITIO N	RESIDUES
1	A.thaliana	1-8, 24-36, 121-141, 194-206	MADSEADK, VNSPNPEVPVTQF, SEGDNSLKDPATKSYAQVFA, DNLFLSKQLGIDW

2	B.rapa	63-85	SEGDNSLK DPASKSYDQV FRPHH
3	S.lycopersicu m	1-9, 30-49, 126-145, 201-210	MANHVAEEK, LDEAAKMEVAPFSHACTLVS, <mark>SEGNS</mark> LKDPASKAYTQVFAP, FTSRDLGTDW
4	G.max	1-8, 22-53, 92-104, 125-136	MTEGNGDK, NVFTDSQSAEAEVKVAPFSHACSLVSPLFGC L, NTVRKGGSHTRNL, EGNSLRDPASKA
5	V.vinifera	26-50, 83- 103, 120- 133	SQTVDIEVAPFSHACSLVSPLFGCL, DHDIEGNC VRKAGSNSRNLLR, ASDDNSLRNPASTA
6	O.sativa	1-9, 26-36, 77-103, 199-207	<mark>MGSSDGDKP</mark> , QQAPGPAMEVG, ISKLPEMVELDIQK GTVRQAGSHTRNM, TSRNLGMDW
7	Z.mays	1-10, 24- 61, 77-89, 127-155, 205-212, 235-258	MNALKHRKAS, NKKGEGKNSRLLPRACQ HPRSASSPKAAAMGSSQADKP, KQQPAVPMDAGAF, SISTLPSMVERDIQTDTVRKPGSHTRNLL, VRSSAP VICYVDDLFTSRNLGIDW
8	P.trichocarpa	1-8, 30-50, 83-102, 121-139, 197-209	MGDLETEKPL, KSIGTLQSVLDKDVERNS VRKGGSHSRNLLRVKRG

Table 6. Protein disorder by the method of beta-aggregation

S.NO	PROTEIN	RESIDUE POSITION	RESIDUES
1	A.thaliana	20-24, 75-82, 108- 115	LAIIV, ISTLVVMM, LDMVKVLF

2	B.rapa	17-24, 131-135	VSTLVVMM, VIAYL,
3	S.lycopersicum	113-120, 192-202	LDMVKVLF, TVILYIDKLFT
4	G.max	113-118, 191-200	MVRVLF, LIQYIDKLFV
5	V.vinifera	188-192	VILYI
6	O.sativa	45-56, 110-121, 190-194	VSVLFGCLGIAF, IDMVKILFEQIL, VILYV
7	Z.mays	65-69, 96-107, 161- 172	IAVSF, VSVLFGCLGIAF, IDMVKVLFEQIL
8	P.trichocarpa	206-217	QSTFYLTWCTII

Ligand Binding residues of the reference protein ACD11 and its orthologues were predicted through RaptorX. The residues involved in forming the binding pockets for the ligands are given in Table 3.

EIS(N-{(2S,3R,4E)-3-hydroxy-1- [(3-O-sulfo-beta-D-galactopyranosyl) oxy]octadec-4-en-2-yl}dodecanamide) binding residues of acd11 and its orthologous protein model obtained from RaptorX server are shown in figure 4. The residues in the binding site pockets are the same for ACD11 and its orthologues. In active site pocket of the reference protein and its orthologues the most prominent and interactive amino acids are Asparagine and Histidine ranging at the same position as shown in the table. 7





64

(a)

(b)



(c)



Figure 5: Ligand binding site and residue prediction of the reference protein and its orthologues retrieved from RaptorX. (a) Arabidopsis thaliana (b) Glycine max (c) Oryza sativa (d) Zea mayz

Table 7. Binding residues and their score

	A.thaliana		G.max		O.sativa		Z.mays	
	60ASP	0.66	74ASP	0.68	69ASP	0.70	120ASP	0.66
Binding residues	97HIS	0.67	104HIS	0.66	99HIS	0.66	150HIS	0.64

And score

CONCLUSION

In the present study we identified, predicted and comprehensively analyzed the accelerated cell death protein ACD11 of Arabidopsis thaliana and its orthologues like *Glycine max, Poplus trichocarpa, Solanum lycopersicum* form dicot plants and *Oryza sativa* and *Zea mays* from monocots across the diverse group of plant species. By taking advantage of the available computational tools, we perform complete analysis of developmental expression, protein-protein interaction, Co-expression, protein structure prediction and determination of intrinsic protein disorder, and finally ligand residues of the selected proteins. It is obvious form the result analysis that ACD11 and its orthologues mostly express in the last developmental stages of the plant i.e. senescence, which is confirming the role of ACD11 in cell death mechanism of the plant and other associated pathways.

From the study of the Co-expression the result shows that some the membrane bounding proteins SNARE, disease resistance and Autophagy related genes are coexpressed with acd11 and their orthologues, which validate again that during the cell death response either due to biotic or abiotic stress the disease resistance and autophagy related genes are also regulated. Form protein-protein interaction it is clear that the interacting partners of the ACD11 and their orthologues from diverse plants species is Enhanced disease resistance protein EDS1and LAZ1, so it is evident from both the results that there is a strong relationship between the co-expressed and the interacting proteins leading to a regulatory pathway of ACD11. Intrinsic disorder protein study predicts the flexibility in protein conformation from which it can be assumed that this protein has the capability of diverse functioning properties. From ligand binding study it is deduced that the common residues in the active site pocket of the protein are ASPRGINE and HISTDINE with high score of average distance and contact of the residue. Future work will elaborate more about the interacting protein partners and co-expressed proteins to determine a common gene-regulatory network of ACD11 and its role in programmed cell death.

REFERENCES

- 1. Airenne, T.T., Kidron, H., Nymalm, Y., Nylund, M., West, G., Mattjus, P., and Salminen, T.A. (2006). Structural evidence for adaptive ligand binding of glycolipid transfer protein. J.Mol.Biol. 355, 224–236.
- 2. Berkey, R., Bendigeri, D., and Xiao, S. (2012). Sphingolipids and plant defense/disease: the "death" connection and beyond. Front Plant Sci, 3, 68.
- 3. Brodersen, P., M. Petersen, H.M. Pike, B. Olszak, S. Skov, N. Odum, L.B. Jorgensen, R.E. Brown, and J. Mundy. (2002.) Knockout of Arabidopsis accelerated-cell-death11 encoding a sphingosine transfer protein causes activation of programmed cell death and defense. *Genes Dev.* 16, 490–502.
- 4. Brodersen, P., Malinovsky, F.G., He'maty, K., Newman, M.A., and Mundy, J. (2005). The role of salicylic acid in the induction of cell death in Arabidopsis acd11. *Plant Physiol.* 138, 1037–1045.
- 5. Brown, R.E., and Mattjus, P. (2007). Glycolipid transfer proteins. *Biochim. Biophys.* Acta 1771, 746–760.
- Chen, M., Cahoon, E.B., Saucedo-Garci'a, M., Plasencia, J., and Gavilanes-Rui'z, M. (2009). Plant sphingolipids: structure, synthesis, function. In Lipids in Photosynthesis: Essential and Regulatory Functions, H. Wada and N. Murata, eds. (New York: Springer Science).77–115.
- 7. Dunker, A. K., Brown, C. J., Lawson, J. D., Iakoucheva, L. M., & Obradović, Z. (2002). Intrinsic disorder and protein function. Biochemistry, *41*(21), 6573-6582.
- Dunker, A. K., Lawson, J. D., Brown, C. J., Williams, R. M., Romero, P., Oh, J. S., & Obradovic, Z. (2001). Intrinsically disordered protein. Journal of molecular graphics and modelling, *19* (1), 26-59.
- 9. Goodstein DM, Shu S, Howson R, Neupane R, Hayes RD, Fazo J, *et al.* .(2012). Phytozome: a comparative platform for green plant genomics. *Nucleic Acids Res*, 40, 1178–1186.

- 10. Greenberg, J. T. & Yao, N (2004). The role and regulation of programmed cell death in plant–pathogen interactions. Cell. Microbiol. 6, 201–211
- Hofius, D., Schultz-Larsen, T., Joensen, J., Tsitsigiannis, D.I., Petersen, N.H., Mattsson, O., Jorgensen, L.B., Jones, J.D., Mundy, J. and Petersen, M. (2009) Autophagic components contribute to hypersensitive cell death in Arabidopsis. Cell, 137, 773–783.
- 12. Källberg, M., Margaryan, G., Wang, S., Ma, J., & Xu, J. (2014). RaptorX server: a resource for template-based protein structure modeling. In *Protein structure prediction*, 17-27. Humana Press, New York, NY.
- Lam, E. (2004). Controlled cell death, plant survival and development. Nat. Rev. Mol. Cell Biol. 5, 305–315.
- 14. Linding, R., Jensen, L. J., Diella, F., Bork, P., Gibson, T. J., & Russell, R. B. (2003). Protein disorder prediction: implications for structural proteomics. *Structure*, *11*(11), 1453-1459.
- Malinovsky, F.G., Brodersen, P., Fiil, B.K., Mckinney, L.V., Thorgrimsen, S., Beck, M., Nielsen, H.B., Pietra, S., Zipfel, C., Robatzek, S., Petersen, M., Hofius, D., and Mundy, J. (2010). Lazarus1, a DUF300 protein, contributes to programmed cell death associated with Arabidopsis acd11 and the hypersensitive response. PLoSONE5, 10. 1371
- Palma, K., Thorgrimsen, S., Malinovsky, F.G., Fiil, B.K., Nielsen, H.B., Brodersen, P., Hofius, D., Petersen, M., and Mundy, J. (2010). Autoimmunity in Arabidopsis acd11 is mediated by epigenetic regulation of an immune receptor. PLoS Pathog. 6, e1001137.
- 17. Paoletti, M., & Clave, C. (2007). The fungus-specific HET domain mediates programmed cell death in Podospora anserina. Eukaryotic Cell, 6(11), 2001-2008.
- Parker, D., Jhala, U. S., Radhakrishnan, I., Yaffe, M. B., Reyes, C., Shulman, A. I., & Montminy, M. (1998). Analysis of an activator: coactivator complex reveals an essential role for secondary structure in transcriptional activation. Molecular cell, 2(3), 353-359.
- Petersen, N.H., Joensen, J., Mckinney, L.V., Brodersen, P., Petersen, M., Hofius, D., and Mundy, J. (2009). Identification of proteins interacting with Arabidopsis ACD11. J.Plant Physiol. 166, pp. 661–666
- 20. Petersen, N.H., McKinney, L.V., Pike, H., Hofius, D., Zakaria, A., Brodersen, P., Petersen, M., Brown, R.E., and Mundy, J. (2008). Human GLTP and mutant forms of ACD11 suppress cell death in the Arabidopsis acd11 mutant. FEBSJ. 275, 4378–4388.
- 21. Pinan-Lucarre B, Paoletti M, Dementhon K, Coulary-Salin B, Clave C (2003). Autophagy is induced during cell death by in compatibility and is essential for

differentiation in the filamentous fungus Podospora anserina. Mol Micro boil 47:321-333

- 22. Plaxco, K. W., & Gross, M. (2001). Unfolded, yes, but random? Never! Nature structural biology, *8*(8), 659-660.
- Simanshu, D.K., Zhai, X., Munch, D., Markham, J.E., Bielawski, J., Bielawska, A., Malinina, L., Molotkovsky, JG., Mundy, J.W., Patel, D.J., Brown, R.E., (2014). Arabidopsis cell death 11, Is Ceramide-1-Phospahte transfer protein and intermediary regulator of pyhtoceramide levels. Cell Rep, 6, 388-399
- Szklarczyk, D., Franceschini, A., Kuhn, M., Simonovic, M., Roth, A., Minguez, P., Doerks, T., Stark, M., Muller, J., Bork, P., Jensen, L.J., von Mering, C. (2011). The STRING database in 2011: functional interaction networks of proteins, globally integrated and scored. Nucleic Acids Res, 39, 561–568.
- 25. Uversky, V. N. (2002). Natively unfolded proteins: a point where biology waits for physics. Protein science, *11*(4), 739-756.
- 26. Wang, J., & Verkhivker, G. M. (2003). Energy landscape theory, funnels, specificity, and optimal criterion of biomolecular binding. Physical review letters, 90(18), 188101.
- Wang, W., Yang, X., Tangchaiburana, S., Ndeh, R., Markham, J.E., Tsegaye, Y., Dunn, T.M., Wang, G.L., Bellizzi, M., Parsons, J.F., etal. (2008). Aninositolphosphorylceramide synthase is involved in regulation of plant programmed cell death associated with defense in Arabidopsis. Plant Cell 20, 3163–3179.